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ENTRY	SESSION

FULL ESTIMATED COST

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FILE 'AGRICOLA' ENTERED AT 12:06:13 ON 15 SEP 2004

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=> microarray and covalent and (coat or immobilization or immobilized)

L1	0 FILE AGRICOLA
L2	12 FILE BIOTECHNO
L3	0 FILE CONFSCI
L4	0 FILE HEALSAFE
L5	0 FILE IMSDRUGCONF
L6	4 FILE LIFESCI
L7	0 FILE MEDICONF
L8	13 FILE PASCAL

TOTAL FOR ALL FILES

L9	29 MICROARRAY AND COVALENT AND (COAT OR IMMOBILIZATION OR IMMOBILIZED)
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=> 19 and (covalent(8A)(immobilized or immobilization or coating))

L10	0 FILE AGRICOLA
L11	3 FILE BIOTECHNO
L12	0 FILE CONFSCI

L13 0 FILE HEALSAFE  
L14 0 FILE IMSDRUGCONF  
L15 1 FILE LIFESCI  
L16 0 FILE MEDICONF  
L17 7 FILE PASCAL

TOTAL FOR ALL FILES

L18 11 L9 AND (COVALENT(8A)(IMMOBILIZED OR IMMOBILIZATION OR COATING))

=> dup rem

ENTER L# LIST OR (END):l18

DUPLICATE IS NOT AVAILABLE IN 'IMSDRUGCONF, MEDICONF'.

ANSWERS FROM THESE FILES WILL BE CONSIDERED UNIQUE

PROCESSING COMPLETED FOR L18

L19 10 DUP REM L18 (1 DUPLICATE REMOVED)

=> d l19 ibib abs total

L19 ANSWER 1 OF 10 PASCAL COPYRIGHT 2004 INIST-CNRS. ALL RIGHTS RESERVED.  
on STN

ACCESSION NUMBER: 2004-0372760 PASCAL

COPYRIGHT NOTICE: Copyright .COPYRGT. 2004 INIST-CNRS. All rights reserved.

TITLE (IN ENGLISH): Carbohydrate chips for studying high-throughput carbohydrate-protein interactions

AUTHOR: PARK Sungjin; LEE Myung-Ryul; PYO Soon-Jin; SHIN Injae

CORPORATE SOURCE: Department of Chemistry, Yonsei University, Seoul 120-749, Korea, Republic of

SOURCE: Journal of the American Chemical Society, (2004), 126(15), 4812-4819

ISSN: 0002-7863 CODEN: JACSAT

DOCUMENT TYPE: Journal

BIBLIOGRAPHIC LEVEL: Analytic

COUNTRY: United States

LANGUAGE: English

NOTE: ref. et notes dissem.

AVAILABILITY: INIST-551, 354000111658010290

AN 2004-0372760 PASCAL

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AB Carbohydrate-protein interactions play important biological roles in living organisms. For the most part, biophysical and biochemical methods have been used for studying these biomolecular interactions. Less attention has been given to the development of high-throughput methods to elucidate recognition events between carbohydrates and proteins. In the current effort to develop a novel high-throughput tool for monitoring carbohydrate-protein interactions, we prepared carbohydrate **microarrays** by immobilizing maleimide-linked carbohydrates on thiol-derivatized glass slides and carried out lectin binding experiments by using these **microarrays**. The results showed that carbohydrates with different structural features selectively bound to the corresponding lectins with relative binding affinities that correlated with those obtained from solution-based assays. In addition, binding affinities of lectins to carbohydrates were also quantitatively analyzed by determining IC<sub>50</sub> values of soluble carbohydrates with the carbohydrate **microarrays**. To fabricate carbohydrate chips that contained more diverse carbohydrate probes, solution-phase parallel and enzymatic glycosylations were performed. Three model disaccharides were in parallel synthesized in solution-phase and used as carbohydrate probes for the fabrication of carbohydrate chips. Three enzymatic glycosylations on glass slides were consecutively performed to generate carbohydrate **microarrays** that contained the complex oligosaccharide, sialyl Le<sup>x</sup>. Overall, these works demonstrated that carbohydrate chips could be efficiently prepared by **covalent immobilization** of maleimide-linked carbohydrates on the thiol-coated glass slides and

applied for the high-throughput analyses of carbohydrate-protein interactions.

L19 ANSWER 2 OF 10 PASCAL COPYRIGHT 2004 INIST-CNRS. ALL RIGHTS RESERVED.  
on STN

ACCESSION NUMBER: 2003-0406556 PASCAL  
COPYRIGHT NOTICE: Copyright .COPYRGT. 2003 INIST-CNRS. All rights reserved.  
TITLE (IN ENGLISH): Optimization of sol-gel formulations and surface treatments for the development of pin-printed protein **microarrays**  
AUTHOR: RUPCICH Nicholas; GOLDSTEIN Aaron; BRENNAN John D.  
CORPORATE SOURCE: Department of Chemistry, McMaster University, Hamilton, Ontario, L8S 4M1, Canada  
SOURCE: Chemistry of materials, (2003), 15(9), 1803-1811, 38 refs.  
ISSN: 0897-4756  
DOCUMENT TYPE: Journal  
BIBLIOGRAPHIC LEVEL: Analytic  
COUNTRY: United States  
LANGUAGE: English  
AVAILABILITY: INIST-21957, 354000118103980060  
AN 2003-0406556 PASCAL  
CP Copyright .COPYRGT. 2003 INIST-CNRS. All rights reserved.  
AB We report on the development and optimization of a sol-gel-based method for the preparation of protein **microarrays** that has the potential to allow pin-spotting of active proteins for high throughput multianalyte biosensing and screening of protein-small molecule interactions. **Microarrays** were printed onto bare and chemically modified surfaces using the commercially available sol-gel precursors tetraethyl orthosilicate and sodium silicate and the newly developed biocompatible sol-gel precursors monosorbitol silane and diglyceryl silane. Parameters such as the type and level of the buffer, the water-to-silane ratio, and the solution pH were also varied to assess the factors that controlled the production of optimal **microarrays**. Such factors included the ability to pin-print without clogging of the pins, the adhesion of the sol-gel spot to the substrate, the dimensions of the microspot, and the stability of both the microspot and the entrapped protein. The microarraying of active antibodies was successfully demonstrated using an optimized combination of parameters, and such arrays were shown to have significantly higher signal-to-background levels than conventional arrays formed by **covalent immobilization** of antibodies on chemically derivatized surfaces.

L19 ANSWER 3 OF 10 BIOTECHNO COPYRIGHT 2004 Elsevier Science B.V. on STN  
DUPLICATE

ACCESSION NUMBER: 2003:36211425 BIOTECHNO  
TITLE: A versatile multi-platform biochip surface attachment chemistry  
AUTHOR: Manning M.; Harvey S.; Galvin P.; Redmond G.  
CORPORATE SOURCE: G. Redmond, Nanotechnology Group, NMRC, Lee Maltings, Ireland.  
E-mail: gredmond@nmrc.ie  
SOURCE: Materials Science and Engineering C, (03 MAR 2003), 23/3 (347-351), 4 reference(s)  
ISSN: 0928-4931  
PUBLISHER ITEM IDENT.: S0928493102002850  
DOCUMENT TYPE: Journal; Article  
COUNTRY: United Kingdom  
LANGUAGE: English  
SUMMARY LANGUAGE: English  
AN 2003:36211425 BIOTECHNO  
AB A versatile DNA spotting and **immobilization** method for

**covalent** attachment of amino-modified probe oligonucleotides in **microarray** format at glass, native silicon dioxide and CVD silicon nitride substrates is reported. Optimal probe spot printing and attachment buffers are identified for each substrate. Relative areal densities of **immobilized** probes as measured by epi-fluorescence microscopy vary with substrate type reflecting differences in surface morphology and chemistry. Target oligonucleotide hybridization occurs at glass and nitride supported probe **microarrays** in an efficient and reproducible manner with excellent measured fluorescence signal-to-background. .COPYRGT. 2002 Elsevier Science B.V. All rights reserved.

L19 ANSWER 4 OF 10 LIFESCI COPYRIGHT 2004 CSA on STN  
ACCESSION NUMBER: 2003:101740 LIFESCI  
TITLE: Impact of surface chemistry and blocking strategies on DNA **microarrays**  
AUTHOR: Taylor, S.; Smith, S.; Windle, B.; Guiseppi-Elie, A.  
CORPORATE SOURCE: Center for Bioelectronics, Biosensors and Biochips,  
Department of Medicinal Chemistry and Department of  
Chemical Engineering, Virginia Commonwealth University, PO  
Box 843038, 601 West Main Street, Richmond, VA 23284-3038,  
USA; E-mail: guiseppi@vcu.edu  
SOURCE: Nucleic Acids Research [Nucleic Acids Res.], (20030000)  
vol. 31, no. 16, e87.  
ISSN: 0305-1048.  
DOCUMENT TYPE: Journal  
FILE SEGMENT: N  
LANGUAGE: English  
SUMMARY LANGUAGE: English

AB The surfaces and **immobilization** chemistries of DNA **microarrays** are the foundation for high quality gene expression data. Four surface modification chemistries, poly-L-lysine (PLL), 3-glycidoxypyriltrimethoxysilane (GPS), DAB- AM-poly(propyleneimine hexadecaamine) dendrimer (DAB) and 3- aminopropyltrimethoxysilane (APS), were evaluated using cDNA and oligonucleotide sub-arrays. Two un-silanized glass surfaces, RCA-cleaned and immersed in Tris- EDTA buffer were also studied. DNA on amine-modified surfaces was fixed by UV (90 mJ/cm super(2)), while DNA on GPS-modified surfaces was **immobilized** by **covalent** coupling. Arrays were blocked with either succinic anhydride (SA), bovine serum albumin (BSA) or left unblocked prior to hybridization with labeled PCR product. Quality factors evaluated were surface affinity for cDNA versus oligonucleotides, spot and background intensity, spotting concentration and blocking chemistry. Contact angle measurements and atomic force microscopy were performed to characterize surface wettability and morphology. The GPS surface exhibited the lowest background intensity regardless of blocking method. Blocking the arrays did not affect raw spot intensity, but affected background intensity on amine surfaces, BSA blocking being the lowest. Oligonucleotides and cDNA on unblocked GPS-modified slides gave the best signal (spot-to-background intensity ratio). Under the conditions evaluated, the unblocked GPS surface along with amine **covalent** coupling was the most appropriate for both cDNA and oligonucleotide **microarrays**.

L19 ANSWER 5 OF 10 PASCAL COPYRIGHT 2004 INIST-CNRS. ALL RIGHTS RESERVED.  
on STN  
ACCESSION NUMBER: 2003-0001164 PASCAL  
TITLE (IN ENGLISH): Protein **microarrays** on ITO surfaces by a  
direct **covalent** attachment scheme  
AUTHOR: NG H. T.; FANG A.; HUANG L.; LI S. F. Y.  
CORPORATE SOURCE: Department of Chemistry National University of  
Singapore, S117543, Singapore, Singapore  
SOURCE: Langmuir, (2002), 18(16), 6324-6329, 33 refs.  
ISSN: 0743-7463  
DOCUMENT TYPE: Journal

BIBLIOGRAPHIC LEVEL: Analytic  
COUNTRY: United States  
LANGUAGE: English  
AVAILABILITY: INIST-20642

AN 2003-0001164 PASCAL

AB We describe the use of an indium tin oxide (ITO) thin film on a solid support serving as an effective generic **immobilization** platform to achieve one-step direct **covalent** attachment of arrayed proteins with good reproducibility and uniformity. Potential functional analyses on these surfaces involving protein-protein and protein-ligand interactions have been demonstrated. We also show that the approach can be adapted, via a photolithography-derived microwell route, to produce high-density protein **microarrays** which typically could accommodate a significantly higher density while occupying a comparatively smaller footprint than that achievable using currently existing high-precision robotic microarrayer systems, suggesting its potential applications in simultaneous parallel biological assays.

L19 ANSWER 6 OF 10 BIOTECHNO COPYRIGHT 2004 Elsevier Science B.V. on STN

ACCESSION NUMBER: 2002:34411501 BIOTECHNO

TITLE: Selective **immobilization** of proteins to self-assembled monolayers presenting active site-directed capture ligands

AUTHOR: Hodneland C.D.; Lee Y.-S.; Min D.-H.; Mrksich M.

CORPORATE SOURCE: M. Mrksich, Department of Chemistry, University of Chicago, Chicago, IL 60637, United States.  
E-mail: mmrksich@midway.uchicago.edu

SOURCE: Proceedings of the National Academy of Sciences of the United States of America, (16 APR 2002), 99/8 (5048-5052), 24 reference(s)

CODEN: PNASA6 ISSN: 0027-8424

DOCUMENT TYPE: Journal; Conference Article

COUNTRY: United States

LANGUAGE: English

SUMMARY LANGUAGE: English

AN 2002:34411501 BIOTECHNO

AB This paper describes a method for the selective and **covalent immobilization** of proteins to surfaces with control over the density and orientation of the protein. The strategy is based on binding of the serine esterase cutinase to a self-assembled monolayer presenting a phosphonate ligand and the subsequent displacement reaction that covalently binds the ligand to the enzyme active site. Surface plasmon resonance (SPR) spectroscopy showed that cutinase binds irreversibly to a monolayer presenting the capture ligand at a density of 1% mixed among tri(ethylene glycol) groups. The **covalent immobilization** is specific for cutinase, and the glycolterminated monolayer effectively prevents unwanted nonspecific adsorption of proteins. To demonstrate that the method could be used to immobilize proteins of interest, a cutinase-calmodulin fusion protein was constructed and **immobilized** to the monolayer. SPR showed that calcineurin selectively associated with the **immobilized** calmodulin. This capture ligand **immobilization** method combines the advantages that the **immobilization** reaction is highly selective for the intended protein, the tether is **covalent** and, hence, stable, and the method avoids the need for synthetic modification and rigorous purification of proteins before **immobilization**. These characteristics make the method well suited to a range of applications and, in particular, for constructing protein **microarrays**.

L19 ANSWER 7 OF 10 PASCAL COPYRIGHT 2004 INIST-CNRS. ALL RIGHTS RESERVED.  
on STN

ACCESSION NUMBER: 2002-0575581 PASCAL

TITLE (IN ENGLISH): Surface characterization of a silicon-chip-based DNA

**microarray**

AUTHOR: LENIGK R.; CARLES M.; IP N. Y.; SUCHER N. J.  
CORPORATE SOURCE: Biotechnology Research Institute Department of Biology  
Hong Kong Univ. of Sci. and Technol., Kowloon, SAR,  
Hong Kong  
SOURCE: Langmuir, (2001), 17(8), 2497-2501, 27 refs.  
ISSN: 0743-7463  
DOCUMENT TYPE: Journal  
BIBLIOGRAPHIC LEVEL: Analytic  
COUNTRY: United States  
LANGUAGE: English  
AVAILABILITY: INIST-20642

AN 2002-0575581 PASCAL

AB The **immobilization** of DNA (deoxyribonucleic acid) on solid supports is a crucial step for any application in the field of DNA **microarrays**. It determines the efficacy of the hybridization and influences the signal strength for the detection. We used solid supports made from silicon wafers as an alternative substrate to the commonly used microscope glass slides. The **covalent immobilization** of thiol-terminated DNA oligonucleotides on self-assembled layers of (3-mercaptopropyl)trimethoxysilane (MPTS) by disulfide bond formation was investigated. Contact angle measurement, variable angle spectral ellipsometry (VASE), X-ray photoelectron spectroscopy (XPS), and atomic force microscopy (AFM) were used to characterize the changing properties of the surface during the DNA array fabrication. During wafer processing the contact angle changed from 3° for the hydroxylated surface to 48.5° after deposition of MPTS. XPS data demonstrated that all sulfur in the MPTS layer was present in the form of reduced SH or S-S groups. VASE measurements indicated a layer thickness of 57.8 Å for the **immobilized** 16 base oligonucleotides including a 18 carbon atom spacer located between the disulfide bond and the oligomer. AFM was used to characterize the DNA layer before and after hybridization to a complementary target. The data recorded after hybridization revealed a sharp increase in particle size from 89 nm<sup>2</sup> to a mean value of 363 nm<sup>2</sup>. Fluorescence microscopy was used to monitor the hybridization of a fluorescently labeled DNA target to the **immobilized** probe. The heat stable disulfide-linkage formed during the oligonucleotide **immobilization** allowed the stripping of complementary DNA targets as well as rehybridization. These data show the advantages and applicability of silicon wafers that have been processed with CMOS (complementary metal oxide semiconductor) compatible processes as solid support in DNA technology. This approach offers the possibility of integration with other silicon-based components such as PCR microreactors and capillary electrophoresis units into a "lab-on-a-chip".

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ACCESSION NUMBER: 2002-0196328 PASCAL

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TITLE (IN ENGLISH): **Covalent** strategy for **immobilization** of DNA-microspots suitable for **microarrays** with label-free and time-resolved optical detection of hybridization  
Miniaturization and chip technology in analytical chemistry

AUTHOR: JUNG A.; STEMMLER I.; BRECHT A.; GAUGLITZ G.  
KITAMORI Takehito (ed.)

CORPORATE SOURCE: Institute of Physical and Theoretical Chemistry,  
University of Tuebingen, Auf der Morgenstelle 8, 72076  
Tuebingen, Germany, Federal Republic of; Cytion SA  
Biopole, Ch. des Croisettes 22, 1066 Epalinges,  
Switzerland  
Department of Applied Chemistry, School of

Engineering, The University of Tokyo, 7-3-1 Hongo,  
Kunkyo-ku, Tokyo 113-8658, Japan

SOURCE: Fresenius' journal of analytical chemistry, (2001),  
371(2), 128-136, 36 refs.  
ISSN: 0937-0633

DOCUMENT TYPE: Journal  
BIBLIOGRAPHIC LEVEL: Analytic  
COUNTRY: Germany, Federal Republic of  
LANGUAGE: English  
AVAILABILITY: INIST-853, 354000096406090060

AN 2002-0196328 PASCAL  
CP Copyright .COPYRGT. 2002 INIST-CNRS. All rights reserved.  
AB Sequence-specific detection and quantification of nucleic acids are central steps in many molecular biology procedures which have also been transferred to chip-based procedures. Hybridization-based assays can be used to quantify and discriminate between DNA target sequences down to the level of single base mismatches. Arrays of DNA probes **immobilized** on a support enable simultaneous testing of multiple sequences of a single sample. DNA arrays can be produced either by in-situ synthesis of oligonucleotides or by **immobilization** of pre-assembled DNA probes. **Covalent** and directed **immobilization** improves the reproducibility and stability of DNA arrays. This is especially interesting with repeated use of transducers or chips. Procedures are described for effective **covalent immobilization** of pre-assembled amino-linked oligonucleotides, by use of ink-jet techniques, on a modified and heated glass surface, with addressable surface areas ranging from 0.01 mm.sup.2 to a few mm.sup.2. Almost immediate evaporation of the spotted droplets on the heated surfaces leads to very high surface hybridization capacities. The surfaces are suitable for use with a label-free detection method - reflectometric interference spectroscopy (RIfS). It is shown that hybridization capacity and non-specific interaction at these DNA-surfaces can be characterized by use of RIfS. With a consumption of less than 80 ng.mm.sup.-.sup.2 oligonucleotide and a specific hybridization capacity of more than 300 fmol mm.sup.-.sup.2, the activated aminodextran procedure was usually suitable for setting up a DNA array with label-free detection. Non-specific interactions with random oligomers or protein (ovalbumin) were low. Up to 150 repeated regenerations (stripping) of the surfaces by acid treatment and denaturing agents, and 50 days of storage, have been possible without significant loss of hybridization capacity.

L19 ANSWER 9 OF 10 BIOTECHNO COPYRIGHT 2004 Elsevier Science B.V. on STN  
ACCESSION NUMBER: 1999:29024605 BIOTECHNO  
TITLE: **Immobilization** of oligonucleotides onto a glass support via disulfide bonds: A method for preparation of DNA **microarrays**  
AUTHOR: Rogers Y.-H.; Jiang-Baucom P.; Huang Z.-J.; Bogdanov V.; Anderson S.; Boyce- Jacino M.T.  
CORPORATE SOURCE: M.T. Boyce-Jacino, Orchid Biocomputer, Inc., Alpha Center, Johns Hopkins Bayview Res. Campus, 5210 Eastern Avenue, Baltimore, MD 21224, United States.  
E-mail: mbj@orchidbio.com  
SOURCE: Analytical Biochemistry, (01 JAN 1999), 266/1 (23-30), 42 reference(s)  
CODEN: ANBCA2 ISSN: 0003-2697  
DOCUMENT TYPE: Journal; Article  
COUNTRY: United States  
LANGUAGE: English  
SUMMARY LANGUAGE: English  
AN 1999:29024605 BIOTECHNO  
AB The **covalent** attachment of disulfide-modified oligonucleotides to a mercaptosilane-modified glass surface is described. This method provides an efficient and specific **covalent** attachment chemistry for **immobilization** of DNA probes onto a solid

support. Glass slides were derivatized with 3- mercaptopropyl silane for attachment of 5-prime disulfide-modified oligonucleotides via disulfide bonds. An attachment density of approximately  $3 \times 10^5$  oligonucleotides/ $\mu\text{m}^2$  was observed. Oligonucleotides attached by this method provided a highly efficient substrate for nucleic acid hybridization and primer extension assays. In addition, we have demonstrated patterning of multiple DNA probes on a glass surface utilizing this attachment chemistry, which allows for array densities of at least 20,000 spots/ $\text{cm}^2$ .

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ACCESSION NUMBER: 1997-0334856 PASCAL  
COPYRIGHT NOTICE: Copyright .COPYRG. 1997 INIST-CNRS. All rights reserved.  
TITLE (IN ENGLISH): Fast temporal response fiber-optic chemical sensors based on the photodeposition of micrometer-scale polymer arrays  
AUTHOR: HEALEY B. G.; WALT D. R.  
CORPORATE SOURCE: The Max Tishler Laboratory for Organic Chemistry, Tufts University, Medford, Massachusetts 02155, United States  
SOURCE: Analytical chemistry : (Washington, DC), (1997), 69(11), 2213-2216, 14 refs.  
ISSN: 0003-2700 CODEN: ANCHAM  
DOCUMENT TYPE: Journal  
BIBLIOGRAPHIC LEVEL: Analytic  
COUNTRY: United States  
LANGUAGE: English  
AVAILABILITY: INIST-120B, 354000061602880400

AN 1997-0334856 PASCAL  
CP Copyright .COPYRG. 1997 INIST-CNRS. All rights reserved.  
AB Fiber-optic chemical sensor **microarrays** for the detection of pH and O.sub.2 have been developed with subsecond response times. Sensor **microarrays** are fabricated by the **covalent immobilization** (pH sensor arrays) or the physical entrapment (O.sub.2 sensor arrays) of fluorescent indicators in photodeposited polymer matrices on optical imaging fibers. Polymer **microarrays** are comprised of thousands of individual elements photodeposited as hemispheres such that each element of the sensor array is coupled directly to a discrete optical element of the imaging fiber and is not in contact with other neighboring elements. Because of the hemispherical shape and the individuality of the array elements, diffusion of analyte to the sensor elements is dominated by radial diffusion, resulting in a rapid response time. pH-sensitive arrays based on fluorescein respond to a 1.5-unit pH change within 300 ms, while the O2-sensitive arrays respond to O.sub.2 changes within 200 ms (90% of steady state response).

=> array and covalent and (coat or immobilization or immobilized)

L20 0 FILE AGRICOLA  
L21 21 FILE BIOTECHNO  
L22 1 FILE CONFSCI  
L23 0 FILE HEALSAFE  
L24 0 FILE IMSDRUGCONF  
L25 8 FILE LIFESCI  
L26 0 FILE MEDICNF  
L27 22 FILE PASCAL

TOTAL FOR ALL FILES

L28 52 ARRAY AND COVALENT AND (COAT OR IMMOBILIZATION OR IMMOBILIZED)

=> (covalent(8A)(immobilized or immobilization or coating)) and l28

L29 0 FILE AGRICOLA



L30 4 FILE BIOTECHNO  
L31 1 FILE CONFSCI  
L32 0 FILE HEALSAFE  
L33 0 FILE IMSDRUGCONF  
L34 3 FILE LIFESCI  
L35 0 FILE MEDICONF  
L36 10 FILE PASCAL

TOTAL FOR ALL FILES

L37 18 (COVALENT(8A) (IMMOBILIZED OR IMMOBILIZATION OR COATING)) AND L28

=> dup rem

ENTER L# LIST OR (END):l37

DUPLICATE IS NOT AVAILABLE IN 'IMSDRUGCONF, MEDICONF'.

ANSWERS FROM THESE FILES WILL BE CONSIDERED UNIQUE

PROCESSING COMPLETED FOR L37

L38 16 DUP REM L37 (2 DUPLICATES REMOVED)

=> d l38 ibib abs total

L38 ANSWER 1 OF 16 PASCAL COPYRIGHT 2004 INIST-CNRS. ALL RIGHTS RESERVED.  
on STN

ACCESSION NUMBER: 2004-0257090 PASCAL

COPYRIGHT NOTICE: Copyright .COPYRG. 2004 INIST-CNRS. All rights reserved.

TITLE (IN ENGLISH): Reversible catalyst supporting via hydrogen-bonding-mediated self-assembly for Atom transfer radical polymerization of MMA

AUTHOR: JUN YANG; SHIJIE DING; RADOSZ Maciej; YOUQING SHEN

CORPORATE SOURCE: Department of Chemical & Petroleum Engineering, University of Wyoming, Laramie, Wyoming 82071, United States

SOURCE: Macromolecules, (2004), 37(5), 1728-1734

ISSN: 0024-9297 CODEN: MAMOBX

DOCUMENT TYPE: Journal

BIBLIOGRAPHIC LEVEL: Analytic

COUNTRY: United States

LANGUAGE: English

NOTE: 1/4 p. ref. et notes

AVAILABILITY: INIST-13789, 354000113490820120

AN 2004-0257090 PASCAL

CP Copyright .COPYRG. 2004 INIST-CNRS. All rights reserved.

AB Atom transfer radical polymerization (ATRP) is a very useful living/controlled radical polymerization process for polymer synthesis, but its products are contaminated with transition metal catalyst residue. Catalyst **immobilization** on solids via **covalent** binding has the advantages of easy catalyst separation and reuse, but it deteriorates the control of the polymerization due to the slowed radical deactivation, which causes chain termination and uncontrolled propagation. In this paper, we report a reversible catalyst supporting concept via hydrogen-bonding-mediated self-assembly. The support acts as a "catalyst sponge" releasing the catalyst as free molecules at elevated temperatures for effective catalysis but absorbing the catalyst after the polymerization for separation. The support was polystyrene gel functionalized with maleimide or thymine units, and the catalyst was tethered on a diaminopyridine unit. A triple hydrogen bond **array** formed between maleimide or thymine and diaminopyridine at room temperature but broke at elevated temperatures. At 60 °C, the reversibly supported catalyst efficiently polymerized MMA in a well-controlled living manner, yielding PMMA with polydispersity as low as those by unsupported catalysts. The recycled catalysts still mediated MMA polymerization with a much improved control.

L38 ANSWER 2 OF 16 PASCAL COPYRIGHT 2004 INIST-CNRS. ALL RIGHTS RESERVED.

on STN

ACCESSION NUMBER: 2004-0087191 PASCAL  
TITLE (IN ENGLISH): The **immobilization** of DNA on microstructured patterns fabricated by maskless lithography  
AUTHOR: ZHANG G. J.; TANII T.; ZAKO T.; FUNATSU T.; OHDOMARI I.  
CORPORATE SOURCE: Nanotechnology Research Center Waseda University, Shinjuku-ku, Tokyo 162-0041, Japan  
SOURCE: Sensors and Actuators, B: Chemical, (2004), 97(2-3), 243-248, 19 refs.  
ISSN: 0925-4005 CODEN: SABCEB  
DOCUMENT TYPE: Journal  
BIBLIOGRAPHIC LEVEL: Analytic  
COUNTRY: Switzerland  
LANGUAGE: English  
AVAILABILITY: INIST-19425 B

AN 2004-0087191 PASCAL

AB The site-directed **covalent immobilization** of amino-terminated DNA oligonucleotides on microstructured patterns at silicon surfaces generated by the methodology of electron beam (EB) lithography was investigated. The microstructured patterns characterized by scanning electron microscopy (SEM) revealed remarkably regular in size and shape. After treatment with different time of activation (10s and 30min), self-assembled layers of 3-aminopropyltriethoxysilane (APTES) on silicon surfaces characterized by X-ray photoelectron spectroscopy (XPS) were demonstrated to obtain similar N 1s peaks. The **immobilization** specificity was evaluated by means of 5\$PRM amino-modified oligonucleotides labeled with Cy 5 at its 3\$PRM end attached onto microstructured patterns. The high-density DNA **array** (40,000 spots per cm<sup>2</sup>) was achieved, and the resulting **array** exhibited the specific binding due to DNA-DNA interaction. Additional studies indicated hardly visible signals when non-complementary probes were **immobilized** on the microstructured patterns. The deposition of DNA in a microstructure **array** using this technique is precise and homogeneous, showing the potential for high-density information storage and the miniaturization for biosensors and biochips. .COPYRG. 2003 Elsevier B.V. All rights reserved.

L38 ANSWER 3 OF 16 PASCAL COPYRIGHT 2004 INIST-CNRS. ALL RIGHTS RESERVED.  
on STN

ACCESSION NUMBER: 2003-0266682 PASCAL  
TITLE (IN ENGLISH): A novel self-assembled nanoparticulate film for **covalent** attachment of antibodies to plastic  
AUTHOR: CUNNINGHAM E.; CAMPBELL C. J.  
CORPORATE SOURCE: Scottish Ctr. Genomic Technol. Info. Univ. of Edinburgh College of Medicine, Edinburgh, EH16 4SB, United Kingdom  
SOURCE: Langmuir, (2003), 19(10), 4509-4511, 12 refs.  
ISSN: 0743-7463 CODEN: LANGD5  
DOCUMENT TYPE: Journal  
BIBLIOGRAPHIC LEVEL: Analytic  
COUNTRY: United States  
LANGUAGE: English  
AVAILABILITY: INIST-20642

AN 2003-0266682 PASCAL

AB Nanostructured silica-layer films were used as solid-phase support for **covalent immobilization** of capture antibody. This silica film showed higher protein loading per cm<sup>2</sup> over conventional two-dimensional surfaces. A sensitive quantitative immunoassay was demonstrated on this silica surface with an optical readout.

L38 ANSWER 4 OF 16 PASCAL COPYRIGHT 2004 INIST-CNRS. ALL RIGHTS RESERVED.  
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ACCESSION NUMBER: 2003-0406556 PASCAL  
 COPYRIGHT NOTICE: Copyright .COPYRGT. 2003 INIST-CNRS. All rights reserved.  
 TITLE (IN ENGLISH): Optimization of sol-gel formulations and surface treatments for the development of pin-printed protein microarrays  
 AUTHOR: RUPCICH Nicholas; GOLDSTEIN Aaron; BRENNAN John D.  
 CORPORATE SOURCE: Department of Chemistry, McMaster University, Hamilton, Ontario, L8S 4M1, Canada  
 SOURCE: Chemistry of materials, (2003), 15(9), 1803-1811, 38 refs.  
 ISSN: 0897-4756  
 DOCUMENT TYPE: Journal  
 BIBLIOGRAPHIC LEVEL: Analytic  
 COUNTRY: United States  
 LANGUAGE: English  
 AVAILABILITY: INIST-21957, 354000118103980060  
 AN 2003-0406556 PASCAL  
 CP Copyright .COPYRGT. 2003 INIST-CNRS. All rights reserved.  
 AB We report on the development and optimization of a sol-gel-based method for the preparation of protein microarrays that has the potential to allow pin-spotting of active proteins for high throughput multianalyte biosensing and screening of protein-small molecule interactions. Microarrays were printed onto bare and chemically modified surfaces using the commercially available sol-gel precursors tetraethyl orthosilicate and sodium silicate and the newly developed biocompatible sol-gel precursors monosorbitol silane and diglyceryl silane. Parameters such as the type and level of the buffer, the water-to-silane ratio, and the solution pH were also varied to assess the factors that controlled the production of optimal microarrays. Such factors included the ability to pin-print without clogging of the pins, the adhesion of the sol-gel spot to the substrate, the dimensions of the microspot, and the stability of both the microspot and the entrapped protein. The microarraying of active antibodies was successfully demonstrated using an optimized combination of parameters, and such **arrays** were shown to have significantly higher signal-to-background levels than conventional **arrays** formed by **covalent immobilization** of antibodies on chemically derivatized surfaces.

L38 ANSWER 5 OF 16 LIFESCI COPYRIGHT 2004 CSA on STN

ACCESSION NUMBER: 2003:101740 LIFESCI  
 TITLE: Impact of surface chemistry and blocking strategies on DNA microarrays  
 AUTHOR: Taylor, S.; Smith, S.; Windle, B.; Guiseppi-Elie, A.  
 CORPORATE SOURCE: Center for Bioelectronics, Biosensors and Biochips, Department of Medicinal Chemistry and Department of Chemical Engineering, Virginia Commonwealth University, PO Box 843038, 601 West Main Street, Richmond, VA 23284-3038, USA; E-mail: guiseppi@vcu.edu  
 SOURCE: Nucleic Acids Research [Nucleic Acids Res.], (20030000) vol. 31, no. 16, e87.  
 ISSN: 0305-1048.  
 DOCUMENT TYPE: Journal  
 FILE SEGMENT: N  
 LANGUAGE: English  
 SUMMARY LANGUAGE: English  
 AB The surfaces and **immobilization** chemistries of DNA microarrays are the foundation for high quality gene expression data. Four surface modification chemistries, poly-L-lysine (PLL), 3-glycidoxypropyltrimethoxysilane (GPS), DAB- AM-poly(propyleminime hexadecaamine) dendrimer (DAB) and 3- aminopropyltrimethoxysilane (APS), were evaluated using cDNA and oligonucleotide sub-**arrays**. Two un-silanized glass surfaces, RCA-cleaned and immersed in Tris- EDTA buffer were also studied. DNA on amine-modified surfaces was fixed by UV (90

mJ/cm super(2)), while DNA on GPS-modified surfaces was **immobilized** by **covalent** coupling. **Arrays** were blocked with either succinic anhydride (SA), bovine serum albumin (BSA) or left unblocked prior to hybridization with labeled PCR product. Quality factors evaluated were surface affinity for cDNA versus oligonucleotides, spot and background intensity, spotting concentration and blocking chemistry. Contact angle measurements and atomic force microscopy were preformed to characterize surface wettability and morphology. The GPS surface exhibited the lowest background intensity regardless of blocking method. Blocking the **arrays** did not affect raw spot intensity, but affected background intensity on amine surfaces, BSA blocking being the lowest. Oligonucleotides and cDNA on unblocked GPS-modified slides gave the best signal (spot-to-background intensity ratio). Under the conditions evaluated, the unblocked GPS surface along with amine **covalent** coupling was the most appropriate for both cDNA and oligonucleotide microarrays.

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ACCESSION NUMBER: 2002-0575581 PASCAL  
TITLE (IN ENGLISH): Surface characterization of a silicon-chip-based DNA microarray  
AUTHOR: LENIGK R.; CARLES M.; IP N. Y.; SUCHER N. J.  
CORPORATE SOURCE: Biotechnology Research Institute Department of Biology  
Hong Kong Univ. of Sci. and Technol., Kowloon, SAR,  
Hong Kong  
SOURCE: Langmuir, (2001), 17(8), 2497-2501, 27 refs.  
ISSN: 0743-7463  
DOCUMENT TYPE: Journal  
BIBLIOGRAPHIC LEVEL: Analytic  
COUNTRY: United States  
LANGUAGE: English  
AVAILABILITY: INIST-20642

AN 2002-0575581 PASCAL

AB The **immobilization** of DNA (deoxyribonucleic acid) on solid supports is a crucial step for any application in the field of DNA microarrays. It determines the efficacy of the hybridization and influences the signal strength for the detection. We used solid supports made from silicon wafers as an alternative substrate to the commonly used microscope glass slides. The **covalent immobilization** of thiol-terminated DNA oligonucleotides on self-assembled layers of (3-mercaptopropyl)trimethoxysilane (MPTS) by disulfide bond formation was investigated. Contact angle measurement, variable angle spectral ellipsometry (VASE), X-ray photoelectron spectroscopy (XPS), and atomic force microscopy (AFM) were used to characterize the changing properties of the surface during the DNA **array** fabrication. During wafer processing the contact angle changed from 3° for the hydroxylated surface to 48.5° after deposition of MPTS. XPS data demonstrated that all sulfur in the MPTS layer was present in the form of reduced SH or S-S groups. VASE measurements indicated a layer thickness of 57.8 Å for the **immobilized** 16 base oligonucleotides including a 18 carbon atom spacer located between the disulfide bond and the oligomer. AFM was used to characterize the DNA layer before and after hybridization to a complementary target. The data recorded after hybridization revealed a sharp increase in particle size from 89 nm<sup>2</sup> to a mean value of 363 nm<sup>2</sup>. Fluorescence microscopy was used to monitor the hybridization of a fluorescently labeled DNA target to the **immobilized** probe. The heat stable disulfide-linkage formed during the oligonucleotide **immobilization** allowed the stripping of complementary DNA targets as well as rehybridization. These data show the advantages and applicability of silicon wafers that have been processed with CMOS (complementary metal oxide semiconductor) compatible processes as solid support in DNA technology. This approach offers the possibility of integration with other silicon-based components such as PCR microreactors

and capillary electrophoresis units into a "lab-on-a-chip".

L38 ANSWER 7 OF 16 PASCAL COPYRIGHT 2004 INIST-CNRS. ALL RIGHTS RESERVED.  
on STN

ACCESSION NUMBER: 2002-0029558 PASCAL  
TITLE (IN ENGLISH): Fabrication of polymer thin films and **arrays**  
with spatial and topographical controls  
AUTHOR: BARTLETT M. A.; YAN M.  
CORPORATE SOURCE: Department of Chemistry Portland State University,  
Portland, OR 97271, United States  
SOURCE: Advanced Materials, (2001), 13(19), 1449-1451, 23  
refs.  
ISSN: 0935-9648 CODEN: ADVMEW  
DOCUMENT TYPE: Journal  
BIBLIOGRAPHIC LEVEL: Analytic  
COUNTRY: Germany, Federal Republic of  
LANGUAGE: English  
AVAILABILITY: INIST-22427

AN 2002-0029558 PASCAL

AB A versatile technique for the **covalent immobilization**  
of polymer thin films on silicon substrates using functionalized  
perfluorophenyl azides (PFPA) was developed. The photochemical  
**immobilization** of poly(2-ethyl-2-oxazoline) PEOX and polystyrene  
thin films was also reported. The ability of the method to create  
spatially defined polymer hybrid **arrays** of differing heights  
resulting in unique surface topographies was demonstrated.

L38 ANSWER 8 OF 16 BIOTECHNO COPYRIGHT 2004 Elsevier Science B.V. on STN  
DUPLICATE

ACCESSION NUMBER: 2001:33043016 BIOTECHNO  
TITLE: Enzyme microgels in packed-bed bioreactors with  
downstream amperometric detection using  
microfabricated interdigitated microsensor electrode  
**arrays**  
AUTHOR: Guiseppi-Elie A.; Sheppard N.F. Jr.; Brahim S.;  
Narinesingh D.  
CORPORATE SOURCE: A. Guiseppi-Elie, Department of Chemical Engineering,  
Center for Bioelectronics, Virginia Commonwealth  
University, P.O. Box 843028, Richmond, VA 23284-3028,  
United States.  
E-mail: guiseppi@vcu.edu  
SOURCE: Biotechnology and Bioengineering, (20 NOV 2001), 75/4  
(475-484), 29 reference(s)  
CODEN: BIBIAU ISSN: 0006-3592  
DOCUMENT TYPE: Journal; Article  
COUNTRY: United States  
LANGUAGE: English  
SUMMARY LANGUAGE: English

AN 2001:33043016 BIOTECHNO

AB In this article, we describe the use of pH-responsive hydrogels as  
matrices for the **immobilization** of two enzymes, glucose oxidase  
(GOx) and glutamate oxidase (GlutOx). Spherical hydrogel beads were  
prepared by inverse suspension polymerization and the enzymes were  
**immobilized** by either physical entrapment or **covalent**  
**immobilization** within or on the hydrogel surface. Packed-bed  
bioreactors were prepared containing the bioactive hydrogels and these  
incorporated into flow injection (FI) systems for the quantitation of  
glucose and monosodium glutamate (MSG) respectively. The FI amperometric  
detector comprised a microfabricated interdigitated **array**  
within a thin-layer flow cell. For the FI manifold incorporating  
**immobilized** GOx, glucose response curves were found to be linear  
over the concentration range 1.8-280 mg dL.sup.-.sup.1 (0.1-15.5 mM) with  
a detection limit of 1.4 mg dL.sup.-.sup.1(0.08 mM). Up to 20 samples can  
be manually analyzed per hour, with the hydrogel-GOx bioreactor

exhibiting good within-day (0.19%) precision. The optimized FI manifold for MSG quantitation yielded a linear response range of up to 135 mg dL<sup>sup.-.sup.1</sup> (8 mM) with a detection limit of 3.38 mg dL<sup>sup.-.sup.1</sup> (0.2 mM) and a throughput of 30 samples h<sup>sup.-.sup.1</sup>. Analysis of commercially produced soup samples gave a within-day precision of 3.6%. Bioreactors containing these two physically entrapped enzymes retained > 60% of their initial activities after a storage period of up to 1 year. .COPYRGT. 2001 John Wiley & Sons, Inc.

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ACCESSION NUMBER: 2002-0196328 PASCAL

COPYRIGHT NOTICE: Copyright .COPYRGT. 2002 INIST-CNRS. All rights reserved.

TITLE (IN ENGLISH): **Covalent** strategy for **immobilization** of DNA-microspots suitable for microarrays with label-free and time-resolved optical detection of hybridization  
Miniaturization and chip technology in analytical chemistry

AUTHOR: JUNG A.; STEMLER I.; BRECHT A.; GAUGLITZ G.  
KITAMORI Takehito (ed.)

CORPORATE SOURCE: Institute of Physical and Theoretical Chemistry,  
University of Tuebingen, Auf der Morgenstelle 8, 72076  
Tuebingen, Germany, Federal Republic of; Cytion SA  
Biopole, Ch. des Croisettes 22, 1066 Epalinges,  
Switzerland

Department of Applied Chemistry, School of  
Engineering, The University of Tokyo, 7-3-1 Hongo,  
Kunkyo-ku, Tokyo 113-8658, Japan  
SOURCE: Fresenius' journal of analytical chemistry, (2001),  
371(2), 128-136, 36 refs.  
ISSN: 0937-0633

DOCUMENT TYPE: Journal

BIBLIOGRAPHIC LEVEL: Analytic

COUNTRY: Germany, Federal Republic of

LANGUAGE: English

AVAILABILITY: INIST-853, 354000096406090060

AN 2002-0196328 PASCAL

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AB Sequence-specific detection and quantification of nucleic acids are central steps in many molecular biology procedures which have also been transferred to chip-based procedures. Hybridization-based assays can be used to quantify and discriminate between DNA target sequences down to the level of single base mismatches. **Arrays** of DNA probes **immobilized** on a support enable simultaneous testing of multiple sequences of a single sample. DNA **arrays** can be produced either by in-situ synthesis of oligonucleotides or by **immobilization** of pre-assembled DNA probes. **Covalent** and directed **immobilization** improves the reproducibility and stability of DNA **arrays**. This is especially interesting with repeated use of transducers or chips. Procedures are described for effective **covalent immobilization** of pre-assembled amino-linked oligonucleotides, by use of ink-jet techniques, on a modified and heated glass surface, with addressable surface areas ranging from 0.01 mm<sup>sup.2</sup> to a few mm<sup>sup.2</sup>. Almost immediate evaporation of the spotted droplets on the heated surfaces leads to very high surface hybridization capacities. The surfaces are suitable for use with a label-free detection method - reflectometric interference spectroscopy (RIfS). It is shown that hybridization capacity and non-specific interaction at these DNA-surfaces can be characterized by use of RIfS. With a consumption of less than 80 ng.mm<sup>sup.-.sup.2</sup> oligonucleotide and a specific hybridization capacity of more than 300 fmol mm<sup>sup.-.sup.2</sup>, the activated aminodextran procedure was usually suitable for setting up a

DNA **array** with label-free detection. Non-specific interactions with random oligomers or protein (ovalbumin) were low. Up to 150 repeated regenerations (stripping) of the surfaces by acid treatment and denaturing agents, and 50 days of storage, have been possible without significant loss of hybridization capacity.

L38 ANSWER 10 OF 16 BIOTECHNO COPYRIGHT 2004 Elsevier Science B.V. on STN

ACCESSION NUMBER: 2000:30219908 BIOTECHNO

TITLE: Integration of layered redox proteins and conductive supports for bioelectronic applications

AUTHOR: Willner I.; Katz E.

CORPORATE SOURCE: Prof. I. Willner, Institute of Chemistry, Hebrew University of Jerusalem, Jerusalem 91904, Israel.  
E-mail: willnea@vms.huji.ac.il

SOURCE: Angewandte Chemie - International Edition, (03 APR 2000), 39/7 (1180-1218)

CODEN: ACIEAY ISSN: 1433-7851

DOCUMENT TYPE: Journal; General Review

COUNTRY: Germany, Federal Republic of

LANGUAGE: English

SUMMARY LANGUAGE: English

AN 2000:30219908 BIOTECHNO

AB Integration of redox enzymes with an electrode support and formation of an electrical contact between the biocatalysts and the electrode is the fundamental subject of bioelectronics and optobioelectronics. This review addresses the recent advances and the scientific progress in electrically contacted, layered enzyme electrodes, and discusses the future applications of the systems in various bioelectronic devices, for example, amperometric biosensors, sensoric **arrays**, logic gates, and optical memories. This review presents the methods for the **immobilization** of redox enzymes on electrodes and discusses the **covalent** linkage of proteins, the use of supramolecular affinity complexes, and the reconstitution of apo-redox enzymes for the nanoengineering of electrodes with protein monolayers of electrodes with protein monolayers and multilayers. Electrical contact in the layered enzyme electrode is achieved by the application of diffusional electron mediators, such as ferrocene derivatives, ferricyanide, quinones, and bipyridinium salts. **Covalent** tethering of electron relay units to layered enzyme electrodes, the cross-linking of affinity complexes formed between redox proteins and electrodes functionalized with relay-cofactor units, or surface reconstitution of apo-enzymes on relay-cofactor-functionalized electrodes yield bioelectrocatalytic electrodes. The application of the functionalized electrodes as biosensor devices is addressed and further application of electrically 'wired' enzymes as catalytic interfaces in biofuel cells is discussed. The organization of sensor **arrays**, self-calibrated biosensors, or gated bioelectronic devices requires the microstructuring of biomaterials on solid supports in the form of ordered micro-patterns. For example, light-sensitive layers composed of azides, benzophenone, or diazine derivatives associated with solid supports can be irradiated through masks to enable the patterned **covalent** linkage of biomaterials to surfaces. Alternatively, patterning of biomaterials can be accomplished by noncovalent interactions (such as in affinity complexes between avidin and a photolabeled biotin, or between an antibody and a photoisomerizable antigen layer) to provide a means of organizing protein microstructures on surfaces. The organization of patterned hydrophilic/hydrophobic domains on surfaces, by using photolithography, stamping, or micromachining methods, allows the selective patterning of surfaces by hydrophobic, noncovalent interactions. Photoactivated layered enzyme electrodes act as light-switchable optobioelectronic systems for the amperometric transduction of recorded photonic information. These systems can act as optical memories, biomolecular amplifiers, or logic gates. The photoswitchable enzyme electrodes are generated by the tethering of photoisomerizable groups to the protein, the reconstitution

of apo-enzymes with semisynthetic photoisomerizable cofactor units, or the coupling of photoisomerizable electron relay units.

L38 ANSWER 11 OF 16 LIFESCI COPYRIGHT 2004 CSA on STN

ACCESSION NUMBER: 2000:90493 LIFESCI

TITLE: Preparation of DNA and protein micro **arrays** on glass slides coated with an agarose film

AUTHOR: Afanassiev, V.; Hanemann, V.; Woelfl, S.

CORPORATE SOURCE: Hans-Knoell-Institut fuer Naturstoff-Forschung, Beutenbergstrasse 11, D-07745 Jena, Germany; E-mail: stefan@imb-jena.de

SOURCE: Nucleic Acids Research [Nucleic Acids Res.], (20000615) vol. 28, no. 12, pp. E66-E66. ISSN: 0305-1048.

DOCUMENT TYPE: Journal

FILE SEGMENT: N

LANGUAGE: English

SUMMARY LANGUAGE: English

AB A thin layered agarose film on microscope slides provides a versatile support for the preparation of arrayed molecular libraries. An activation step leading to the formation of aldehyde groups in the agarose creates reactive sites that allow **covalent immobilization** of molecules containing amino groups. **Arrays** of oligonucleotides and PCR products were prepared by tip printing. After hybridization with complementary fluorescence labeled nucleic acid probes strong fluorescence signals of sequence-specific binding to the **immobilized** probes were detected. The intensity of the fluorescence signals was proportional to the relative amount of **immobilized** oligonucleotides and to the concentration of the fluorescence labeled probe. We also used the agarose film-coated slides for the preparation of protein **arrays**. In combination with specific fluorescence labeled antibodies these protein **arrays** can be used for fluorescence linked immune assays. With this approach different protein tests can be performed in parallel in a single reaction with minimal amounts of the binding reagents.

L38 ANSWER 12 OF 16 PASCAL COPYRIGHT 2004 INIST-CNRS. ALL RIGHTS RESERVED. on STN

ACCESSION NUMBER: 1999-0360277 PASCAL

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TITLE (IN ENGLISH): Design of oligonucleotide **arrays** at interfaces

AUTHOR: BONCHEVA M.; SCHEIBLER L.; LINCOLN P.; VOGEL H.; AKERMAN B.

CORPORATE SOURCE: Department of Physical Chemistry, Chalmers University of Technology, 412 96 Goeteborg, Sweden; Institut de Chimie Organique, Universite de Lausanne, 1015 Lausanne, Switzerland; Laboratoire de Chimie Physique des Polymeres et Membranes, Ecole Polytechnique Federale de Lausanne, 1015 Lausanne, Switzerland

SOURCE: Langmuir, (1999), 15(13), 4317-4320, 15 refs.

ISSN: 0743-7463 CODEN: LANGD5

DOCUMENT TYPE: Journal

BIBLIOGRAPHIC LEVEL: Analytic

COUNTRY: United States

LANGUAGE: English

AVAILABILITY: INIST-20642, 354000085516580030

AN 1999-0360277 PASCAL

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AB The surface attachment and detection of DNA probes are essential in the design of nucleic acid-based biosensors. A new strategy for the **covalent immobilization** of single-stranded oligonucleotides on gold-covered planar supports is presented.



Optimization of the surface density in the resulting DNA **arrays** permits a high hybridization efficiency to be achieved. Surface plasmon resonance and, for the first time, ATR-FTIR spectroscopy are used to follow in situ the oligonucleotide layer formation and the subsequent complementary strand hybridization. Such well-defined, covalently **immobilized** oligonucleotide **arrays** can find application in the development of novel DNA-based sensors for mutation detection and gene mapping as well as in studies of nucleic acid-ligand interactions.

L38 ANSWER 13 OF 16 BIOTECHNO COPYRIGHT 2004 Elsevier Science B.V. on STN  
 ACCESSION NUMBER: 1999:29221649 BIOTECHNO  
 TITLE: A novel microtiter plate based method for identification of B-cell epitopes  
 AUTHOR: Gregorius K.; Dalum I.; Freisleben M.; Mouritsen S.; Elsnér H.I.  
 CORPORATE SOURCE: K. Gregorius, M and E Biotech A-S, 6 Kogle Alle, DK-2970 Horsholm, Denmark.  
 SOURCE: Journal of Peptide Science, (1999), 5/2 (75-82), 21 reference(s)  
 CODEN: JPSIEI ISSN: 1075-2617  
 DOCUMENT TYPE: Journal; Article  
 COUNTRY: United Kingdom  
 LANGUAGE: English  
 SUMMARY LANGUAGE: English  
 AN 1999:29221649 BIOTECHNO  
 AB A new type of microtiter plate capable of binding biomolecules covalently in a one step procedure was used to map linear B-cell epitopes in two different proteins using a peptide-based solid phase immunoassay. The method was compared with a conventional **immobilization** method using passive adsorption to microtiter plates. An **array** of 15-mer peptides, overlapping by five amino acids, representing the entire sequences of ubiquitin and murine tumor necrosis factor- $\alpha$ , respectively, was synthesized. The peptides were **immobilized** covalently using the new, specialized microtiter plates or non-covalently using conventional ELISA microtiter plates of the high binder type. Subsequently, specific antisera to ubiquitin or murine tumor necrosis factor- $\alpha$  were added to identify potential linear B-cell epitopes. All peptides, which were recognized on the conventional microtiter plates, were also recognized on the plates with the covalently bound peptides. In addition, the **covalent immobilization** method revealed epitopes that were not identified using the method for non-**covalent** binding although the peptides were in fact present on the non-**covalent** binding surface. The interaction with the hydrophobic surface of the conventional microtiter plate apparently interfered negatively with antibody recognition. The covalently binding microtiter plates described here could be useful for identification of new B-cell epitopes in protein antigens.

L38 ANSWER 14 OF 16 BIOTECHNO COPYRIGHT 2004 Elsevier Science B.V. on STN  
 ACCESSION NUMBER: 1999:29024605 BIOTECHNO  
 TITLE: **Immobilization** of oligonucleotides onto a glass support via disulfide bonds: A method for preparation of DNA microarrays  
 AUTHOR: Rogers Y.-H.; Jiang-Baucom P.; Huang Z.-J.; Bogdanov V.; Anderson S.; Boyce- Jacino M.T.  
 CORPORATE SOURCE: M.T. Boyce-Jacino, Orchid Biocomputer, Inc., Alpha Center, Johns Hopkins Bayview Res. Campus, 5210 Eastern Avenue, Baltimore, MD 21224, United States. E-mail: mbj@orchidbio.com  
 SOURCE: Analytical Biochemistry, (01 JAN 1999), 266/1 (23-30), 42 reference(s)  
 CODEN: ANBCA2 ISSN: 0003-2697  
 DOCUMENT TYPE: Journal; Article  
 COUNTRY: United States

LANGUAGE: English

SUMMARY LANGUAGE: English

AN 1999:29024605 BIOTECHNO

AB The **covalent** attachment of disulfide-modified oligonucleotides to a mercaptosilane-modified glass surface is described. This method provides an efficient and specific **covalent** attachment chemistry for **immobilization** of DNA probes onto a solid support. Glass slides were derivatized with 3- mercaptopropyl silane for attachment of 5-prime disulfide-modified oligonucleotides via disulfide bonds. An attachment density of approximately  $3 \times 10^{5.5}$  oligonucleotides/ $\mu\text{m}^{2.2}$  was observed. Oligonucleotides attached by this method provided a highly efficient substrate for nucleic acid hybridization and primer extension assays. In addition, we have demonstrated patterning of multiple DNA probes on a glass surface utilizing this attachment chemistry, which allows for **array** densities of at least 20,000 spots/ $\text{cm}^{2.2}$ .

L38 ANSWER 15 OF 16 PASCAL COPYRIGHT 2004 INIST-CNRS. ALL RIGHTS RESERVED.  
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ACCESSION NUMBER: 1997-0334856 PASCAL

COPYRIGHT NOTICE: Copyright .COPYRGT. 1997 INIST-CNRS. All rights reserved.

TITLE (IN ENGLISH): Fast temporal response fiber-optic chemical sensors based on the photodeposition of micrometer-scale polymer **arrays**

AUTHOR: HEALEY B. G.; WALT D. R.

CORPORATE SOURCE: The Max Tishler Laboratory for Organic Chemistry, Tufts University, Medford, Massachusetts 02155, United States

SOURCE: Analytical chemistry : (Washington, DC), (1997), 69(11), 2213-2216, 14 refs.

ISSN: 0003-2700 CODEN: ANCHAM

DOCUMENT TYPE: Journal

BIBLIOGRAPHIC LEVEL: Analytic

COUNTRY: United States

LANGUAGE: English

AVAILABILITY: INIST-120B, 354000061602880400

AN 1997-0334856 PASCAL

CP Copyright .COPYRGT. 1997 INIST-CNRS. All rights reserved.

AB Fiber-optic chemical sensor microarrays for the detection of pH and  $0.2$  have been developed with subsecond response times. Sensor microarrays are fabricated by the **covalent immobilization** (pH sensor **arrays**) or the physical entrapment ( $0.2$  sensor **arrays**) of fluorescent indicators in photodeposited polymer matrices on optical imaging fibers. Polymer microarrays are comprised of thousands of individual elements photodeposited as hemispheres such that each element of the sensor **array** is coupled directly to a discrete optical element of the imaging fiber and is not in contact with other neighboring elements. Because of the hemispherical shape and the individuality of the **array** elements, diffusion of analyte to the sensor elements is dominated by radial diffusion, resulting in a rapid response time. pH-sensitive **arrays** based on fluorescein respond to a 1.5-unit pH change within 300 ms, while the  $0.2$ -sensitive **arrays** respond to  $0.2$  charges within 200 ms (90% of steady state response).

L38 ANSWER 16 OF 16 CONFSCI COPYRIGHT 2004 CSA on STN

ACCESSION NUMBER: 1998:58409 CONFSCI

DOCUMENT NUMBER: 98-058409

TITLE: Non-manual sequential functionalization of individual electrodes of an **array** for **covalent immobilization** of different biological recognition elements

AUTHOR: Kurzawa, C.; Habermueller, K.; Strohmeier, J.; Schuhmann,

W.  
 CORPORATE SOURCE: Ruhr Universitaet Bochum, Germany  
 SOURCE: Biosensors & Bioelectronics, Institute of BioScience &  
 Technology, Cranfield University, Cranfield, Beds MK43 0AL,  
 United Kingdom; fax: +44 1234 752 401; URL:  
<http://www.elsevier.nl:80/homepage/sah/bios98>, Abstracts  
 and full papers available..  
 Meeting Info.: 982 5025: 5th World Congress on Biosensors  
 (9825025). Berlin (Germany). 3-5 Jun 1998. Institute of  
 BioScience & Technology.  
 DOCUMENT TYPE: Conference  
 FILE SEGMENT: DCCP  
 LANGUAGE: English

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=> (covalent(8A)(immobilized or immobilization or coating)) and array

L39	65 FILE CAPLUS
L40	9 FILE BIOSIS
L41	13 FILE MEDLINE
L42	4 FILE EMBASE
L43	3531 FILE USPATFULL

TOTAL FOR ALL FILES

L44	3622 (COVALENT(8A)(IMMOBILIZED OR IMMOBILIZATION OR COATING)) AND ARRAY
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PROCESSING COMPLETED FOR L40

PROCESSING COMPLETED FOR L41

PROCESSING COMPLETED FOR L42

L45 71 DUP REM L39-L42 (20 DUPLICATES REMOVED)

=> l45 and py<2001

L46 64 S L45  
L47 20 FILE CAPLUS  
L48 3 S L45  
L49 1 FILE BIOSIS  
L50 4 S L45  
L51 0 FILE MEDLINE  
L52 0 S L45  
L53 0 FILE EMBASE  
L54 0 S L45  
L55 0 FILE USPATFULL

TOTAL FOR ALL FILES

L56 21 L45 AND PY<2001

=> d l56 ibib abs total

L56 ANSWER 1 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 2002:276458 CAPLUS  
DOCUMENT NUMBER: 136:274240  
TITLE: Methods and apparatus for nucleic acid analysis  
INVENTOR(S): Drmanac, Radoje  
PATENT ASSIGNEE(S): USA  
SOURCE: U.S. Pat. Appl. Publ., 37 pp., Cont.-in-part of U.S.  
6,297,006.

CODEN: USXXCO

DOCUMENT TYPE: Patent

LANGUAGE: English

FAMILY ACC. NUM. COUNT: 7

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
US 2002042048	A1	20020411	US 1997-892503	19970714
US 6309824	B1	20011030	US 1997-784747	19970116
US 6297006	B1	20011002	US 1997-812951	19970304
US 6383742	B1	20020507	US 1997-912885	19970815
US 2002034737	A1	20020321	US 1997-947779	19971009
WO 9831836	A1	19980723	WO 1998-US704	19980114 <--
W: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, GW, HU, ID, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, UZ, VN, YU, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
RW: GH, GM, KE, LS, MW, SD, SZ, UG, ZW, AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG				
AU 9861317	A1	19980807	AU 1998-61317	19980114 <--
AU 745201	B2	20020314		
EP 968305	A1	20000105	EP 1998-905956	19980114 <--
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI				
JP 2001509027	T2	20010710	JP 1998-534497	19980114
BR 9806914	A	20010918	BR 1998-6914	19980114
NZ 513913	A	20010928	NZ 1998-513913	19980114
US 2003108897	A1	20030612	US 2002-187251	20020701
PRIORITY APPLN. INFO.:				US 1997-784747 A2 19970116
				US 1997-812951 A2 19970304
				US 1997-892503 A2 19970714
				US 1997-912885 A2 19970815

US 1997-947779 B1 19971009  
WO 1998-US704 W 19980114

AB The present invention provides a method for detecting a target nucleic acid species including the steps of providing an **array** of probes affixed to a substrate and a plurality of labeled probes. Each labeled probe is selected to have a first nucleic acid sequence which is complementary to a first portion of a target nucleic acid and wherein the nucleic acid sequence of at least one probe affixed to the substrate is complementary to a second portion of the nucleic acid sequence of the target. The invention relates to applying a target nucleic acid to the **array** under suitable conditions for hybridization of probe sequences to complementary sequences. The method further involves introduction of labeled probe to the **array**, hybridizing a probe affixed to the substrate to the target nucleic acid, hybridizing the labeled probe to the target nucleic acid, affixing the labeled probe to an adjacently hybridized probe in the **array** and detecting the labeled probe affixed to the probe in the **array**. The invention further relates to **covalent** joining of the **immobilized** probes to labeled probes that are immediately adjacent to the immobilized probe on the target sequence; removing any non-ligated labeled probes; detecting the presence of the target nucleic acid by detecting the presence of said labeled probe attached to the immobilized probes.

L56 ANSWER 2 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 2000:900852 CAPLUS  
DOCUMENT NUMBER: 134:53462  
TITLE: Microarrays of immobilized oligonucleotide probes and computer-based systems for gene expression analysis  
INVENTOR(S): Baidya, Narayan; Chen, Yii-Der Ida; Holding, Julie; Yu, Yie-Teh  
PATENT ASSIGNEE(S): Clingenix, Inc., USA  
SOURCE: PCT Int. Appl., 58 pp.  
CODEN: PIXXD2  
DOCUMENT TYPE: Patent  
LANGUAGE: English  
FAMILY ACC. NUM. COUNT: 1  
PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
WO 2000077257	A1	20001221	WO 2000-US15850	20000609 <--
W:	AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM			
RW:	GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG			
EP 1185701	A1	20020313	EP 2000-939721	20000609
R:	AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO			
US 6716579	B1	20040406	US 2000-591366	20000609
PRIORITY APPLN. INFO.:			US 1999-138690P	P 19990611
			WO 2000-US15850	W 20000609

AB The present invention provides microarrays comprising a plurality of polynucleotide probes having sequences complementary to the 3' untranslated region of a gene transcript, whose chromosomal location has been defined. The probes are **immobilized** via **covalent** linkage on a solid support, such as nitrocellulose, nylon, polypropylene, glass, and silicon. Probes comprising sequence tagged site (STS) tags may be used. Polynucleotides conjugated with an enzyme, radioactive, or luminescent label may be contained in the **array**. The

microarrays are particularly useful for conducting comparative gene expression analyses, eg., differential expression of multiple genes. The present invention also includes a method of preparing these microarrays and various methods of using these microarrays for detecting differential expression for multiple gene transcripts amongst multiple subjects. Further provided by the invention are computer readable media recorded thereon an **array** of polynucleotide probes as specified herein, a computer-based system, and kits for detecting differential expression of a multiplicity of gene transcripts. Floppy disks, hard disk, magnetic tape, CD-ROM, random access memory (RAM), or read only memory (ROM).

REFERENCE COUNT: 10 THERE ARE 10 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L56 ANSWER 3 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 2000:682505 CAPLUS

DOCUMENT NUMBER: 134:53237

TITLE: Covalent attachment of oligodeoxyribonucleotides to amine-modified Si (001) surfaces

AUTHOR(S): Strother, Todd; Hamers, Robert J.; Smith, Lloyd M.

CORPORATE SOURCE: Department of Chemistry, University of Wisconsin, Madison, WI, 53706-1396, USA

SOURCE: Nucleic Acids Research (2000), 28(18), 3535-3541

CODEN: NARHAD; ISSN: 0305-1048

PUBLISHER: Oxford University Press

DOCUMENT TYPE: Journal

LANGUAGE: English

AB A recently described reaction for the UV-mediated attachment of alkenes to silicon surfaces is utilized as the basis for the preparation of functionalized silicon surfaces. UV light mediates the reaction of t-butyloxycarbonyl (t-BOC) protected  $\omega$ -unsatd. amino-alkane (10-aminodec-1-ene) with hydrogen terminated silicon (001). Removal of the t-BOC protecting group yields an aminodecane-modified silicon surface. The resultant amino groups can be coupled to thiol-modified oligodeoxyribonucleotides using a heterobifunctional crosslinker, permitting the preparation of DNA **arrays**. Two methods for controlling the surface d. of oligodeoxyribonucleotides were explored: in the first, binary mixts. of 10-aminodec-1-ene and dodecene were utilized in the initial UV-mediated coupling reaction; a linear relationship was found between the mole fraction of aminodecene and the d. of DNA hybridization sites. In the second, only a portion of the t-BOC protecting groups was removed from the surface by limiting the time allowed for the deprotection reaction. The oligodeoxyribonucleotide-modified surfaces were extremely stable and performed well in DNA hybridization assays. These surfaces provide an alternative to gold or glass for surface immobilization of oligonucleotides in DNA **arrays** as well as a route for the coupling of nucleic acid biomol. recognition elements to semiconductor materials.

REFERENCE COUNT: 55 THERE ARE 55 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L56 ANSWER 4 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 2000:636926 CAPLUS

DOCUMENT NUMBER: 133:345235

TITLE: Preparation of DNA and protein micro **arrays** on glass slides coated with an agarose film

AUTHOR(S): Afanassiev, Victor; Hanemann, Vera; Wolf, Stefan

CORPORATE SOURCE: Hans-Knoll-Institut fur Naturstoff-Forschung, Jena, D-07745, Germany

SOURCE: Nucleic Acids Research (2000), 28(12), e66, ii-v

CODEN: NARHAD; ISSN: 0305-1048

PUBLISHER: Oxford University Press

DOCUMENT TYPE: Journal

LANGUAGE: English

AB A thin layered agarose film on microscope slides provides a versatile support for the preparation of arrayed mol. libraries. An activation step leading to the formation of aldehyde groups in the agarose creates reactive sites that allow **covalent immobilization** of mols. containing amino groups. **Arrays** of oligonucleotides and PCR products were prepared by tip printing. After hybridization with complementary fluorescence labeled nucleic acid probes strong fluorescence signals of sequence-specific binding to the immobilized probes were detected. The intensity of the fluorescence signals was proportional to the relative amount of immobilized oligonucleotides and to the concentration of the fluorescence labeled probe. We also used the agarose film-coated slides for the preparation of protein **arrays**. In combination with specific fluorescence labeled antibodies these protein **arrays** can be used for fluorescence linked immune assays. With this approach different protein tests can be performed in parallel in a single reaction with minimal amts. of the binding reagents.

REFERENCE COUNT: 21 THERE ARE 21 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L56 ANSWER 5 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 2000:401982 CAPLUS

DOCUMENT NUMBER: 133:13393

TITLE: Method for efficiently immobilizing oligonucleotide on a carrier via a covalent bond

INVENTOR(S): Ueda, Minoru; Okamoto, Sachiko; Ozaki, Aya; Mineno, Junichi; Kimizuka, Fusao; Asada, Kiyozo; Kato, Ikunoshin

PATENT ASSIGNEE(S): Takara Shuzo Co., Ltd., Japan

SOURCE: PCT Int. Appl., 39 pp.

CODEN: PIXXD2

DOCUMENT TYPE: Patent

LANGUAGE: Japanese

FAMILY ACC. NUM. COUNT: 2

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
WO 2000034457	A1	20000615	WO 1999-JP6867	19991208 <--
W:	AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM			
RW:	GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW, AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG			
EP 1138762	A1	20011004	EP 1999-959692	19991208
R:	AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, SI, LT, LV, FI, RO			
PRIORITY APPLN. INFO.:			JP 1998-351276	A 19981210
			WO 1999-JP6867	W 19991208

AB A method is described for efficiently immobilizing an oligonucleotide on a carrier (e.g., glass, quartz) via a covalent bond (between an amino group introduced in an oligonucleotide and an aldehyde group held on a carrier) with a spacer by spotting a buffer (e.g., morpholine buffer, carbonate buffer) containing the oligonucleotide onto the carrier. A method is also claimed for detecting a target nucleic acid by hybridization using the immobilized oligonucleotide prepared by this method.

REFERENCE COUNT: 6 THERE ARE 6 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L56 ANSWER 6 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 2000:258571 CAPLUS  
DOCUMENT NUMBER: 133:14211  
TITLE: Integration of layered redox proteins and conductive supports for bioelectronic applications  
AUTHOR(S): Willner, Itamar; Katz, Eugenii  
CORPORATE SOURCE: Institute of Chemistry, The Hebrew University of Jerusalem, Jerusalem, 91904, Israel  
SOURCE: Angewandte Chemie, International Edition (2000), 39(7), 1181-1218  
CODEN: ACIEF5; ISSN: 1433-7851  
PUBLISHER: Wiley-VCH Verlag GmbH  
DOCUMENT TYPE: Journal; General Review  
LANGUAGE: English

AB Integration of redox enzymes with an electrode support and formation of an elec. contact between the biocatalysts and the electrode is the fundamental subject of bioelectronics and optobioelectronics. This review, with 254 refs., addresses the recent advances and the scientific progress in elec. contacted, layered enzyme electrodes, and discusses the future applications of the systems in various bioelectronic devices, for example, amperometric biosensors, sensoric **arrays**, logic gates, and optical memories. This review presents the methods for the **immobilization** of redox enzymes on electrodes and discusses the **covalent** linkage of proteins, the use of supramol. affinity complexes, and the reconstitution of apo-redox enzymes for the nanoengineering of electrodes with protein monolayers of electrodes with protein monolayers and multilayers. Elec. contact in the layered enzyme electrode is achieved by the application of diffusional electron mediators, such as ferrocene derivs., ferricyanide, quinones, and bipyridinium salts. Covalent tethering of electron relay units to layered enzyme electrodes, the crosslinking of affinity complexes formed between redox proteins and electrodes functionalized with relay-cofactor units, or surface reconstitution of apo-enzymes on relay-cofactor-functionalized electrodes yield bioelectrocatalytic electrodes. The application of the functionalized electrodes as biosensor devices is addressed and further application of elec. "wired" enzymes as catalytic interfaces in biofuel cells is discussed. The organization of sensor **arrays**, self-calibrated biosensors, or gated bioelectronic devices requires the microstructuring of biomaterials on solid supports in the form of ordered micro-patterns. For example, light-sensitive layers composed of azides, benzophenone, or diazine derivs. associated with solid supports can be irradiated through masks to enable the patterned covalent linkage of biomaterials to surfaces. Alternatively, patterning of biomaterials can be accomplished by noncovalent interactions (such as in affinity complexes between avidin and a photolabeled biotin, or between an antibody and a photoisomerizable antigen layer) to provide a means of organizing protein microstructures on surfaces. The organization of patterned hydrophilic/hydrophobic domains on surfaces, by using photolithog., stamping, or micromachining methods, allows the selective patterning of surfaces by hydrophobic, noncovalent interactions. Photoactivated layered enzyme electrodes act as light-switchable optobioelectronic systems for the amperometric transduction of recorded photonic information. These systems can act as optical memories, biomol. amplifiers, or logic gates. The photoswitchable enzyme electrodes are generated by the tethering of photoisomerizable groups to the protein, the reconstitution of apo-enzymes with semisynthetic photoisomerizable cofactor units, or the coupling of photoisomerizable electron relay units.

REFERENCE COUNT: 74 THERE ARE 74 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L56 ANSWER 7 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 2000:225030 CAPLUS  
DOCUMENT NUMBER: 133:172660  
TITLE: Comparison between Different Strategies of Covalent



Attachment of DNA to Glass Surfaces to Build DNA  
Microarrays

AUTHOR(S): Zammattéo, Nathalie; Jeanmart, Laurent; Hamels,  
Sandrine; Courtois, Stéphane; Louette, Pierre; Hevesi,  
Laszlo; Remacle, Jose  
CORPORATE SOURCE: Laboratoire de Biochimie Cellulaire, Facultés  
Universitaires N.-D. de la Paix, Namur, 5000, Belg.  
SOURCE: Analytical Biochemistry (2000), 280(1),  
143-150  
CODEN: ANBCA2; ISSN: 0003-2697  
PUBLISHER: Academic Press  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB DNA microarray is a powerful tool allowing simultaneous detection of many  
different target mols. present in a sample. The efficiency of the  
**array** depends mainly on the sequence of the capture probes and the  
way they are attached to the support. The coupling procedure must be  
quick, covalent, and reproducible in order to be compatible with automatic  
spotting devices dispensing tiny drops of liqs. on the surface. We  
compared several coupling strategies currently used to covalently graft  
DNA onto a glass surface. The results indicate that fixation of aminated  
DNA to an aldehyde-modified surface is a choice method to build DNA  
microarrays. Both the coupling procedure and the hybridization efficiency  
have been optimized. The detection limit of human cytomegalovirus target  
DNA amplicons on such DNA microarrays has been estimated to be 0.01 nM by  
fluorescent detection. (c) 2000 Academic Press.

REFERENCE COUNT: 24 THERE ARE 24 CITED REFERENCES AVAILABLE FOR THIS  
RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L56 ANSWER 8 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 2000:194696 CAPLUS

DOCUMENT NUMBER: 133:14218

TITLE: Combined atomic force microscopy (AFM), X-ray  
photoelectron spectroscopy (XPS) and quartz crystal  
microbalance (QCM) studies of glucose oxidase (GOx)  
immobilised onto self-assembled monolayer on the gold  
film

AUTHOR(S): Losic, Dusan; Gooding, J. Justin; Shapter, Joe;  
Erokin, Paul; Short, Ken

CORPORATE SOURCE: Faculty of Science and Engineering, The Flinders  
University of South Australia, Adelaide, 5001,  
Australia

SOURCE: Proceedings - Australian Conference on Nuclear  
Techniques of Analysis (1999), 11th, 213-217  
CODEN: PCTAD7; ISSN: 0156-3602

PUBLISHER: Australian Institute of Nuclear Science and  
Engineering

DOCUMENT TYPE: Journal

LANGUAGE: English

AB In fabrication of biosensors, self-assembled monolayers (SAM) are an  
attractive method of immobilizing enzymes at electrode surface since it  
allows precise control over the amount and spatial distribution of the  
immobilized enzyme. The covalent attachment of glucose oxidase (GOx) to a  
carboxylic terminated SAM chemisorbed onto gold films was achieved via  
carbodiimide activation of the carboxylic acids to a reactive intermediate  
susceptible to nucleophilic attack by amines on free lysine chains of the  
enzyme. Atomic force microscopy (AFM), XPS and quartz crystal microbalance  
(QCM) measurements were used for characterization of GOx modified gold  
surfaces. Tapping mode AFM studies have revealed that GOx mols. form  
slightly disordered **arrays** of pentagonal or hexagonal clusters.  
Observed features of immobilized GOx are distributed as a sub-monolayer on  
the SAM surface which has allowed visualization of native and unfolded  
enzyme structure. The presence of the SAM and enzyme on the gold surface  
was detected by XPS spectroscopy. Spectra show typical peaks for the C

1s, O 1s and N 1s regions. A kinetic study of the adsorption of GOx onto activated SAM using in-situ QCM allowed determination the amount of immobilized GOx on the layer and consequently the optimal immobilization conditions. Performance parameters of the biosensor such as sensitivity to glucose concentration as a function of enzyme loading were evaluated amperometrically using the redox mediator p-benzoquinone.

REFERENCE COUNT: 13 THERE ARE 13 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L56 ANSWER 9 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 2000:140565 CAPLUS

DOCUMENT NUMBER: 132:176585

TITLE: Covalent attachment of nucleic acid molecules onto solid phases via disulfide bonds

INVENTOR(S): Anderson, Steve; Rogers, Yu-Hui

PATENT ASSIGNEE(S): Orchid Biocomputer, Inc., USA

SOURCE: U.S., 14 pp., Cont.-in-part of U.S. 5,837,860.

CODEN: USXXAM

DOCUMENT TYPE: Patent

LANGUAGE: English

FAMILY ACC. NUM. COUNT: 2

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
US 6030782	A	20000229	US 1997-975862	19971121 <--
US 5837860	A	19981117	US 1997-812010	19970305 <--
PRIORITY APPLN. INFO.:			US 1997-812010	A2 19970305

AB Nucleic acid mols. are immobilized reversibly onto solid-phases with reversible disulfide bonds for nucleic acid mol. **array** preparation A solid phase surface is coated with mercaptosilane HS-L-Si(Y)(Z)X [L = (CH<sub>2</sub>)<sub>n</sub>, (-CH<sub>2</sub>)<sub>n</sub>-aromatic-(CH<sub>2</sub>)<sub>n</sub>-, or aromatic group (n ≥ 1)); X = alkoxy, acyloxy, halo; Y and Z = alkoxy, acyloxy, halo, or nonhydrolyzable inert group] or disulfidesilane RS-S-L-Si(Y)(Z)X (L, X, Y, Z as above; R = nonhydrolyzable group), which is coupled to sulfhydryl- or disulfide-modified nucleic acid mols. via disulfide bonds. These methods can be used to prepare reusable nucleic acid mol. **arrays** with high specificity and high efficiency. Glass slides are etched and then treated with HS(CH<sub>2</sub>)<sub>3</sub>Si(OMe)<sub>3</sub> in an acidic buffer in aqueous EtOH, cured, and dried. The cured slides are treated with 5'-disulfide modified oligonucleotides in a carbonate buffer to produce a disulfide bond between the oligonucleotide and the silane layer (the disulfide exchange reaction). GBA (Genetic Bit Anal.) primers having a poly-T spacer arm are immobilized by the disulfide exchange reaction onto glass slides for the typing of single nucleotide polymorphisms, the immediately 3'-distal sequences of which are complementary to the primers.

REFERENCE COUNT: 14 THERE ARE 14 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L56 ANSWER 10 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 2000:31381 CAPLUS

DOCUMENT NUMBER: 132:74513

TITLE: Covalent attachment of oligonucleotide probes to derivatized polypropylene supports

INVENTOR(S): Rampal, Jang B.

PATENT ASSIGNEE(S): Beckman Coulter, Inc., USA

SOURCE: U.S., 18 pp.

CODEN: USXXAM

DOCUMENT TYPE: Patent

LANGUAGE: English

FAMILY ACC. NUM. COUNT: 1

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
US 6013789	A	20000111	US 1998-26742	19980220 <--
PRIORITY APPLN. INFO.:			US 1998-26742	19980220

AB Disclosed herein is a method for attaching pre-synthesized oligonucleotides to a polypropylene support medium. Most preferably, a polypropylene film is aminated by a plasma discharge in the presence of ammonia gas. An oligonucleotide having a terminal phosphate is activated in the presence of an imidazole (N-methylimidazole or 4,5-dicyanoimidazole) and a carbodiimide (EDC, 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide) to form a phosphorimidazolidine. The activated oligonucleotide becomes immobilized by forming a phosphoramidate bond with the aminated polypropylene. The invention can be used to construct oligonucleotide **arrays** for hybridization assays.

REFERENCE COUNT: 9 THERE ARE 9 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L56 ANSWER 11 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 1999:331955 CAPLUS  
DOCUMENT NUMBER: 131:194952  
TITLE: Design of Oligonucleotide **Arrays** at Interfaces  
AUTHOR(S): Boncheva, Mila; Scheibler, Lukas; Lincoln, Per; Vogel, Horst; Aakerman, Bjoern  
CORPORATE SOURCE: Department of Physical Chemistry, Chalmers University of Technology, Goeteborg, S-412 96, Swed.  
SOURCE: Langmuir (1999), 15(13), 4317-4320  
CODEN: LANGD5; ISSN: 0743-7463  
PUBLISHER: American Chemical Society  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB The surface attachment and detection of DNA probes are essential in the design of nucleic acid-based biosensors. A new strategy for the **covalent immobilization** of single-stranded oligonucleotides on gold-covered planar supports is presented. Optimization of the surface d. in the resulting DNA **arrays** permits a high hybridization efficiency to be achieved. Surface plasmon resonance and, for the first time, ATR-FTIR spectroscopy are used to follow in situ the oligonucleotide layer formation and the subsequent complementary strand hybridization. Such well-defined, covalently immobilized oligonucleotide **arrays** can find application in the development of novel DNA-based sensors for mutation detection and gene mapping as well as in studies of nucleic acid-ligand interactions.

REFERENCE COUNT: 20 THERE ARE 20 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L56 ANSWER 12 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 1999:309643 CAPLUS  
DOCUMENT NUMBER: 131:126035  
TITLE: Versatile derivatization of solid support media for covalent bonding on DNA-microchips  
AUTHOR(S): Beier, Markus; Hoheisel, Jorg D.  
CORPORATE SOURCE: Functional Genome Analysis, Deutsches Krebsforschungszentrum, Heidelberg, D-69120, Germany  
SOURCE: Nucleic Acids Research (1999), 27(9), 1970-1977  
CODEN: NARHAD; ISSN: 0305-1048  
PUBLISHER: Oxford University Press  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB A chemical was developed that permits on DNA-**arrays** both the **covalent immobilization** of pre-fabricated nucleic acids-such as oligonucleotides, PCR-products or peptide nucleic acid oligomers-and the in situ synthesis of such compds. on either glass or

polypropylene surfaces. Bonding was found to be stable even after some 30 cycles of stripping. Due to a dendrimeric structure of the linker mol., the loading can be modified in a controlled manner and increased beyond the capacity of glass without neg. effects on hybridization efficiency. Also, the chemical warrants the modulation of other surface properties such as charge or hydrophobicity. Preferentially, attachment of nucleic acids takes place only via the terminal amino-group of amino-modified oligonucleotides or the terminal hydroxyl-group of unmodified mols. so that the entire mol. is accessible to probe hybridization. This derivatization represents a support chemical versatile enough to serve nearly all current forms of DNA-arrays or microchips.

REFERENCE COUNT: 23 THERE ARE 23 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L56 ANSWER 13 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 1999:220206 CAPLUS

DOCUMENT NUMBER: 130:249124

TITLE: Method for the detection and isolation of biomolecules via molecular recognition using immobilized pyranosyl nucleotide supramolecular structures

INVENTOR(S): Windhab, Norbert; Miculka, Christian; Hoppe, Hans-Ullrich

PATENT ASSIGNEE(S): Hoechst A.-G., Germany

SOURCE: Ger. Offen., 14 pp.

CODEN: GWXXBX

DOCUMENT TYPE: Patent

LANGUAGE: German

FAMILY ACC. NUM. COUNT: 1

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
DE 19741716	A1	19990325	DE 1997-19741716	19970922 <--
CA 2303086	AA	19990401	CA 1998-2303086	19980921 <--
WO 9915893	A1	19990401	WO 1998-EP6001	19980921 <--
W: AU, BR, CA, JP, KR, US				
RW: AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE				
AU 9913340	A1	19990412	AU 1999-13340	19980921 <--
AU 757912	B2	20030313		
EP 1018007	A1	20000712	EP 1998-956830	19980921 <--
R: AT, BE, CH, FR, GB, IT, LI, NL, SE				
BR 9812490	A	20000926	BR 1998-12490	19980921 <--
JP 2001517795	T2	20011009	JP 2000-513140	19980921
PRIORITY APPLN. INFO.:			DE 1997-19741716	A 19970922
			WO 1998-EP6001	W 19980921

AB The invention concerns biosensors for the detection of mols. that are composed of an **array** of **immobilized** supramol. structures that have non-covalent binding sites for the receptor mols.; the receptor mols. are selected in a manner that they recognize the target mols. and they are labeled; the receptors recognize the target mols. immunol.; the receptor mol. with the captured target is hybridized to the immobilized **array** of the biosensor. The hybridized complex can be detected by the sensor in various ways, e.g. by fluorescence, change in electrode potential etc. Changing thermodyn. parameters, e.g. concentration, temperature, the hybridized receptor-target complex can be removed from the surface and used in further procedures. The immobilized binding mols. are nucleic acid derivs. that differ from those in biol. samples; e.g. pyranosyl nucleotides, and pyranosyl RNAs; the immobilized binding mols. can hybridize several types of receptor mols. The receptor mols. are chosen from chemical libraries, peptide libraries; several receptor mols. that recognize different parts of the target mols. are used; the recognizing peptide/protein part of the receptor is bound

via linkers to the hybridizing part. The method can be used in combination with biochip techniques for drug screening, pesticide and herbicide research, for anal. and production of catalysts.

L56 ANSWER 14 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 1999:187354 CAPLUS  
DOCUMENT NUMBER: 130:350879  
TITLE: A novel microtiter plate based method for  
identification of B-cell epitopes  
AUTHOR(S): Gregorius, Klaus; Dalum, Iben; Freisleben, Marianne;  
Mouritsen, Soren; Elsner, Henrik I.  
CORPORATE SOURCE: M and E Biotech A/S, Horsholm, DK-2970, Den.  
SOURCE: Journal of Peptide Science (1999), 5(2),  
75-82  
CODEN: JPSIEI; ISSN: 1075-2617  
PUBLISHER: John Wiley & Sons Ltd.  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB A new type of microtiter plate capable of binding biomols. covalently in a one step procedure was used to map linear B-cell epitopes in two different proteins using a peptide-based solid phase immunoassay. The method was compared with a conventional immobilization method using passive adsorption to microtiter plates. An **array** of 15-mer peptides, overlapping by five amino acids, representing the entire sequences of ubiquitin and murine tumor necrosis factor- $\alpha$ , resp., was synthesized. The peptides were immobilized covalently using the new, specialized microtiter plates or non-covalently using conventional ELISA microtiter plates of the high binder type. Subsequently, specific antisera to ubiquitin or murine tumor necrosis factor- $\alpha$  were added to identify potential linear B-cell epitopes. All peptides, which were recognized on the conventional microtiter plates, were also recognized on the plates with the covalently bound peptides. In addition, the **covalent immobilization** method revealed epitopes that were not identified using the method for non-covalent binding although the peptides were in fact present on the non-covalent binding surface. The interaction with the hydrophobic surface of the conventional microtiter plate apparently interfered neg. with antibody recognition. The covalently binding microtiter plates described here could be useful for identification of new B-cell epitopes in protein antigens.

REFERENCE COUNT: 21 THERE ARE 21 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L56 ANSWER 15 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 1999:24966 CAPLUS  
DOCUMENT NUMBER: 130:205645  
TITLE: Immobilization of oligonucleotides onto a glass  
support via disulfide bonds: a method for preparation  
of DNA microarrays  
AUTHOR(S): Rogers, Yu-Hui; Jiang-Baucom, Ping; Huang, Zhi-Jian;  
Bogdanov, Valery; Anderson, Stephen; Boyce-Jacino,  
Michael T.  
CORPORATE SOURCE: Alpha Center, Orchid Biocomputer, Inc., Baltimore, MD,  
21224, USA  
SOURCE: Analytical Biochemistry (1999), 266(1),  
23-30  
CODEN: ANBCA2; ISSN: 0003-2697  
PUBLISHER: Academic Press  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB The covalent attachment of disulfide-modified oligonucleotides to a mercaptosilane-modified glass surface is described. This method provides an efficient and specific **covalent** attachment chemical for **immobilization** of DNA probes onto a solid support. Glass slides were derivatized with 3-mercaptopropyl silane for attachment of 5-prime

disulfide-modified oligonucleotides via disulfide bonds. An attachment d. of approx.  $3 \times 10^5$  oligonucleotides/ $\mu\text{m}^2$  was observed. Oligonucleotides attached by this method provided a highly efficient substrate for nucleic acid hybridization and primer extension assays. In addition, we have demonstrated patterning of multiple DNA probes on a glass surface utilizing this attachment chemical, which allows for **array** densities of at least 20,000 spots/ $\text{cm}^2$ . (c) 1999 Academic Press.

REFERENCE COUNT: 42 THERE ARE 42 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L56 ANSWER 16 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 1998:618855 CAPLUS

DOCUMENT NUMBER: 129:226631

TITLE: Covalent attachment of nucleic acid molecules onto solid phases via disulfide bonds

INVENTOR(S): Anderson, Stephen; Rogers, Yu-hui

PATENT ASSIGNEE(S): Molecular Tool, Inc., USA

SOURCE: PCT Int. Appl., 40 pp.

CODEN: PIXXD2

DOCUMENT TYPE: Patent

LANGUAGE: English

FAMILY ACC. NUM. COUNT: 2

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
WO 9839481	A1	19980911	WO 1998-US4114	19980304 <--
W: AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GE, GH, GM, GW, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, AM, AZ, BY, KG, KZ, MD, RU, TJ, TM				
RW: GH, GM, KE, LS, MW, SD, SZ, UG, ZW, AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG				
US 5837860	A	19981117	US 1997-812010	19970305 <--
CA 2252911	AA	19980911	CA 1998-2252911	19980304 <--
AU 9865402	A1	19980922	AU 1998-65402	19980304 <--
AU 739010	B2	20011004		
EP 910670	A1	19990428	EP 1998-911457	19980304 <--
R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT, IE, FI				
JP 2000510710	T2	20000822	JP 1998-538691	19980304 <--
PRIORITY APPLN. INFO.: US 1997-812010 A 19970305				
WO 1998-US4114 W 19980304				

OTHER SOURCE(S): MARPAT 129:226631

AB Nucleic acid mols. are immobilized reversibly onto solid-phases with reversible disulfide bonds for nucleic acid mol. **array** preparation. A solid phase surface is coated with mercaptosilane, e.g., HS-L-Si(Y)(Z)X [L = (CH<sub>2</sub>)<sub>n</sub>, (-CH<sub>2</sub>)<sub>n</sub>-aromatic-(CH<sub>2</sub>)<sub>n</sub>-, or aromatic group (n .gtorsim. 1)); X = alkoxy, acyloxy, halo; Y and Z = alkoxy, acyloxy, halo, or nonhydrolyzable inert group], which is coupled to sulfhydryl- or disulfide-modified nucleic acid mol. via a disulfide bond. These methods can be used to prepare reusable nucleic acid mol. **arrays** with high specificity and high efficiency. Glass slides are etched and then treated with HS(CH<sub>2</sub>)<sub>3</sub>Si(OMe)<sub>3</sub> in an acidic buffer in aqueous EtOH, cured, and dried. The cured slides are treated with 5'-disulfide modified oligonucleotides in a carbonate buffer to produce a disulfide bond between the oligonucleotide and the silane layer (the disulfide exchange reaction). GBA (Genetic Bit Anal.) primers having a poly-T spacer arm are immobilized by the disulfide exchange reaction onto glass slides for the typing of single nucleotide polymorphisms, the immediately 3'-distal sequences of which are complementary to the primers.

REFERENCE COUNT: 7 THERE ARE 7 CITED REFERENCES AVAILABLE FOR THIS

L56 ANSWER 17 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 1997:283725 CAPLUS

DOCUMENT NUMBER: 126:324555

TITLE: Fast Temporal Response Fiber-Optic Chemical Sensors  
Based on the Photodeposition of Micrometer-Scale  
Polymer **Arrays**

AUTHOR(S): Healey, Brian G.; Walt, David R.

CORPORATE SOURCE: Max Tishler Laboratory for Organic Chemistry, Tufts  
University, Medford, MA, 02155, USA

SOURCE: Analytical Chemistry (1997), 69(11),  
2213-2216

CODEN: ANCHAM; ISSN: 0003-2700

PUBLISHER: American Chemical Society

DOCUMENT TYPE: Journal

LANGUAGE: English

AB Fiber-optic chemical sensor microarrays for the detection of pH and O<sub>2</sub> were developed with subsecond response times. Sensor microarrays are fabricated by the **covalent immobilization** (pH sensor **arrays**) or the phys. entrapment (O<sub>2</sub> sensor **arrays**) of fluorescent indicators in photodeposited polymer matrixes on optical imaging fibers. Polymer microarrays are comprised of thousands of individual elements photodeposited as hemispheres such that each element of the sensor **array** is coupled directly to a discrete optical element of the imaging fiber and is not in contact with other neighboring elements. Because of the hemispherical shape and the individuality of the **array** elements, diffusion of analyte to the sensor elements is dominated by radial diffusion, resulting in a rapid response time. PH-sensitive **arrays** based on fluorescein respond to a 1.5-unit pH change within 300 ms, while the O<sub>2</sub>-sensitive **arrays** respond to O<sub>2</sub> changes within 200 ms (90% of steady state response).

L56 ANSWER 18 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 1996:487411 CAPLUS

DOCUMENT NUMBER: 125:189895

TITLE: Surface **arrays** of energy absorbing polymers  
enabling covalent attachment of biomolecules for  
subsequent laser-induced uncoupling/desorption

AUTHOR(S): Voivodov, Kamen I.; Ching, Jesus; Hutchens, T. William

CORPORATE SOURCE: Mol. Anal. Syst., Houston, TX, 77056, USA

SOURCE: Tetrahedron Letters (1996), 37(32),  
5669-5672

CODEN: TELEAY; ISSN: 0040-4039

PUBLISHER: Elsevier

DOCUMENT TYPE: Journal

LANGUAGE: English

AB Synthetic polymers with desirable film-forming characteristics were chemical modified to incorporate, covalently, UV energy-absorbing mols. (EAM) of a type used for matrix-assisted laser desorption/ionization (MALDI) time-of-flight mass spectrometry. The polymer-bound EAM were activated to covalently bind macromols., e.g., peptides, through the carboxyl group of the bound EAM. The EAM was shown to act as a photolabile macromol. tether. Biomols. covalently bound to surface **arrays** of EAM-polymer were uncoupled and desorbed/ionized with single pulses of laser irradiation

L56 ANSWER 19 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 1996:217236 CAPLUS

TITLE: Site-directed mutagenesis of enzymes to facilitate  
controlled immobilization for bioreactors and  
biosensors

AUTHOR(S): Huang, Wei; Bachas, Leonidas G.; Bhattacharyya,  
Dibakar

CORPORATE SOURCE: Department Chemistry, University Kentucky, Lexington,  
KY, 40506, USA  
SOURCE: Book of Abstracts, 211th ACS National Meeting, New  
Orleans, LA, March 24-28 (1996), BIOT-080.  
American Chemical Society: Washington, D. C.  
CODEN: 62PIAJ

DOCUMENT TYPE: Conference; Meeting Abstract  
LANGUAGE: English

AB **Immobilization** of enzymes by **covalent** attachment has many applications in the development of biosensors and bioreactors. Conventional chemical immobilization methods often result in random orientation of enzyme mols. on the immobilization surface and in a substantial decrease of the enzymic activity. In this presentation, the modification of enzymes by site-directed mutagenesis to facilitate their controlled immobilization will be described. The cysteine-free proteinase subtilisin was chosen as a model protein to demonstrate the feasibility of this approach. A single cysteine residue was introduced at a position away from the active site of the enzyme by site-directed mutagenesis. The genetically modified enzyme was then immobilized on several surfaces through the -SH group on the cysteine. Compared to the conventional random immobilization method, site-directed immobilization yields controlled two-dimensional enzyme **arrays** with a higher specific activity on the surface.

L56 ANSWER 20 OF 21 CAPLUS COPYRIGHT 2004 ACS on STN

ACCESSION NUMBER: 1993:555125 CAPLUS  
DOCUMENT NUMBER: 119:155125  
TITLE: An amperometric glucose sensor based on isoporous  
crystalline protein membranes as immobilization matrix  
AUTHOR(S): Neubauer, A.; Pum, D.; Sleytr, U. B.  
CORPORATE SOURCE: Zent. Ultrastrukturforsch., Univ. Bodenkult., Vienna,  
A-1180, Austria  
SOURCE: Analytical Letters (1993), 26(7), 1347-60  
CODEN: ANALBP; ISSN: 0003-2719  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB S-layer ultrafiltration membranes (SUMs) with an active filtration layer composed of coherent two-dimensional, isoporous protein crystals (S-layers) have been used as matrix for immobilizing monolayers of enzymes. Since S-layers are formed by periodic repetition of identical protein subunits, functional groups are present on the crystalline **array** in an identical position and orientation. As a consequence monolayers of enzymes can bind in a geometrically well defined way. For the **covalent immobilization** of enzymes carboxyl groups from the S-layer protein were activated with carbodiimide and allowed to react with amino groups of the enzyme. SUMs were employed as a new type of immobilization matrix for the development of an amperometric glucose sensor using glucose oxidase (GOD) as the biol. active component. Glucose oxidase covalently bound to the surface of the S-layer protein retained approx. 40% of its activity. The enzyme loaded SUMs were covered with a layer of gold or platinum to function as working electrodes. These sensors yielded high signals (150nA/mm<sup>2</sup>/mmol glucose), fast response times (1-30s) and a linearity range up to 12 mM glucose. The stability under working conditions was more than 48 h. There was no loss in activity after a storage period of 6 mo.

L56 ANSWER 21 OF 21 BIOSIS COPYRIGHT (c) 2004 The Thomson Corporation. on STN

ACCESSION NUMBER: 2000:226063 BIOSIS  
DOCUMENT NUMBER: PREV200000226063  
TITLE: Electrostatically driven immobilization of peptides onto  
(maleic anhydride-alt-methyl vinyl ether) copolymers in  
aqueous media.  
AUTHOR(S): Ladaviere, Catherine; Lorenzo, Carmen; Elaissari,



Abdelhamid; Mandrand, Bernard; Delair, Thierry [Reprint author]

CORPORATE SOURCE: Unite Mixte UMR-103, CNRS-bioMerieux, ENS-Lyon, 46, allée d'Italie, 69364, Lyon, France

SOURCE: Bioconjugate Chemistry, (March-April, 2000) Vol. 11, No. 2, pp. 146-152. print.

CODEN: BCCHES. ISSN: 1043-1802.

DOCUMENT TYPE: Article

LANGUAGE: English

ENTRY DATE: Entered STN: 7 Jun 2000

Last Updated on STN: 5 Jan 2002

AB The **covalent immobilization** of a model peptide onto the MAMVE copolymer, via the formation of amide bonds, occurred in moderate yields in aqueous conditions. The improvement of the grafting reaction was achieved by adding at the amino terminus of the model peptide a sequence (tag) of three positively charged amino acids, lysine or arginine, and by taking profit of electrostatic attractive interactions between the negatively charged copolymer and the tagged peptides. The arginine tag was more efficient than the lysine tag for enhancing the immobilization reaction, proving that the effect was due to an electrostatic driving force. On the basis of these results, a tentative mechanism is discussed, and Scatchard plots pointed out two regimes of binding. With the first, at low polymer load (up to 50% of saturation for a lysine tag and 60-70% for an arginine tag), the binding occurred with a positive cooperative effect, the already bound peptide participating to the binding of others. A second one for higher coverages, for which the binding occurred with a negative cooperativity, and saturation was reached in the presence of a large excess of peptide.